

# Gas-liquid transfer data used to analyze hydrophobic hydration and find the nature of the Kauzmann-Tanford hydrophobic factor

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**Hydrophobic free energy for protein folding is currently measured by liquid-liquid transfer, based on an analogy between the folding process and the transfer of a nonpolar solute from water into a reference solvent. The second part of the analogy (transfer into a nonaqueous solvent) is dubious and has been justified by arguing that transfer out of water probably contributes the major part of the free energy change. This assumption is wrong: transfer out of water contributes no more than half the total, often less. Liquid-liquid transfer of the solute from water to liquid alkane is written here as the sum of 2 gas-liquid transfers: (i) out of water into vapor, and (ii) from vapor into liquid alkane. Both gas-liquid transfers have known free energy values for several alkane solutes. The comparable values of the two different transfer reactions are explained by the values, determined in 1991 for three alkane solutes, of the cavity work and the solute-solvent interaction energy. The transfer free energy is the difference between the positive cavity work and the negative solute-solvent interaction energy. The interaction energy has similar values in water and liquid alkane that are intermediate in magnitude between the cavity work in water and in liquid alkane. These properties explain why the transfer free energy has comparable values (with opposite signs) in the two transfers. The current hydrophobic free energy is puzzling and poorly defined and needs a new definition and method of measurement.**

solvation free energy | hydrophobicity | protein energetics | Ostwald coefficient | Pratt-Chandler analysis

Ever since Kauzmann's revolutionary proposal (1) in 1959, the hydrophobic factor has been widely acknowledged as a major factor in protein folding energetics (1–3). In the widely used Kauzmann-Tanford approach to measuring it, hydrophobic free energy is synonymous with hydrophobic hydration and is measured by the preference of a nonpolar solute for a reference solvent over water, based on the solute's relative solubility in the two solvents. This approach is still the standard one, although it has been criticized. Much of the current research into the nature of the hydrophobic factor is based on computer simulations (4). The basic criticism of the liquid-liquid transfer approach is that gas-liquid transfer (which is better understood) is used to determine the solvation free energy of the nonpolar solute. Moreover, the choice of a reference solvent ought not to change the apparent hydrophobic free energy, but it does (5). The purpose here is to investigate these criticisms with the aid of gas-liquid transfer results and their interpretation.

The 1971 paper by Nozaki and Tanford (2) is often cited as the starting point of quantitative work on measuring hydrophobic hydration. However, as ref. 2 emphasizes, it was based on the protocol developed in the Cohn and Edsall laboratory at Harvard by McMeekin, et al. (6) who used it to investigate the water solubility of polar as well as nonpolar protein side chains. The latter authors found that nonpolar groups have additive  $\Delta G_{\text{hyd}}$  values and consequently that model compound results can be used to interpret this aspect of protein folding energetics.

The protocol chosen by Nozaki and Tanford (2) contains three steps: first, measure the solubility ( $S$ ) (pH 7) of the solute (chosen to model a side chain) in water ( $w$ ) and in a reference solvent ( $r$ ). Crystalline amino acids were used and results with  $r =$  ethanol or dioxane were compared (2). Second, compute  $K_D = (S_r)/(S_w)$  and  $\Delta G_{\text{LL}} = -RT \ln K_D$ . Third, focus on the side chain properties by setting  $\Delta G_{\text{LL}} = \Delta G_{\text{aa}} - \Delta G_{\text{gly}}$  (where aa = amino acid and gly = glycine); subtracting the  $\Delta G_{\text{LL}}$  value for Gly was intended to remove the effects of the  $\alpha\text{-NH}_3^+$  and  $\alpha\text{-COO}^-$  groups. Nozaki and Tanford gave hydrophobicity values for twelve nonpolar protein side chains and they did not study polar side chains.

## The Polar Group Effect

Problems arise if the solute being transferred contains both polar and nonpolar groups. The perturbing polar group effect was discovered in 1935 when McMeekin, et al. (6) found that the characteristic hydrophobic contribution of the  $-\text{CH}_2-$  group is missing when the side chains of aspartic acid, asparagine, glutamic acid, and glutamine are analyzed, apparently because there are nearby polar groups. The polar group effect can be understood by the iceberg model of Frank and Evans (7) and Kauzmann (1, 8). In the iceberg model, the hydrophobic free energy arises chiefly from the perturbed H-bonded structure of water that develops around nonpolar groups (1, 7, 8). Using the iceberg model, the polar group effect is explained as the result of strong interactions between water and polar groups that affect the perturbed water structure around nearby nonpolar groups.

## Criticisms by Hildebrand (1968) and Ben-Naim (1979)

In 1968 Hildebrand (9) criticized the concept of "hydrophobic bonds" [Kauzmann's term (1)] because hydrocarbons interact favorably with water. An oil drop added to a beaker of water does not remain a droplet, but instead spreads out to form a surface film in order to interact with as much water as possible. Tanford pondered his reply for eleven years. In 1979 he agreed (10) that hydrocarbons make favorable van der Waals (dispersion force) interactions with water. He also used data for the interfacial energies of water and hexane to argue, however, that the hydrophobicity of water is caused by its high cohesive energy density, which tends to squeeze out all solutes. Polar solutes resist being squeezed out by making strong interactions with water. Other workers had a different view of the role of the solute-solvent interaction: Sharp, Nicholls, Friedman and Honig said in 1991: "For a nonpolar solute in water, the solute-solvent interaction is the origin of the hydrophobic effect." (11).

Today there are mainly two opposing views (12) on the hydrophobicity of water: either it is caused chiefly by the small size of the water molecule (13) or by the perturbed H-bonded water structure around nonpolar groups (the iceberg model) (1, 7, 8).

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**Table 1. Transfer energetics for gas-liquid transfer of three alkane solutes to water and to liquid alkane**

Solute	Water			Liquid alkane		
	$\Delta G_c$	$E_a$	$\Delta G_{GL}$	$\Delta G_c$	$E_a$	$\Delta G_{GL}$
Propane	8.6	-6.6	2.0	4.3	-6.4	-2.1
Isobutane	10.1	-7.8	2.3	5.8	-8.4	-2.6
Neopentane	11.1	-8.6	2.5	6.6	-9.5	-2.9

Values from (14) are given in kcal/mol at 25 °C.  $\Delta G_c$  is the work of making a cavity for the solute in the solvent;  $E_a$  is the solute-solvent interaction energy;  $\Delta G_{GL}$  is the algebraic sum of  $\Delta G_c$  and  $E_a$ .

are midway between the  $\Delta G_c$  values for water and liquid alkane; this ensures that the magnitudes of the two  $\Delta G_{GL}$  values are comparable. When Kauzmann (1959) and Nozaki and Tanford (1971) wrote their papers (1, 2), the values of  $\Delta G_c$  and  $E_a$  [obtained by Lee (14) in 1991] were not yet available and it wasn't possible to foresee this behavior. The basic result today is that the hydrophobic factor defined by Kauzmann (1) and Nozaki and Tanford (2) is doubtful because it is based on an assumption that is no longer justified. It will be necessary in future work to reassess the hydrophobic free energy found by liquid-liquid transfer. For other current problems involving the hydrophobic factor, see review (26).

The hallmark of the hydrophobic factor in protein folding energetics is its unusual temperature dependence (8), expressed as a large value of  $\Delta Cp$ , the difference between the heat capacity values of the native and unfolded protein forms. The enthalpy change for unfolding increases as much as sixfold between 25 and 110 °C for some proteins (27) and this behavior is attributed entirely to the hydrophobic factor, since the much smaller dependence from the peptide groups has the opposite sign and the enthalpy change from breaking van der Waals contact interactions is considered temperature-independent (28). The unusual temperature dependence of the hydrophobic factor is observed in both model compound results and protein unfolding data (29, 30).

**Other Approaches to Measuring Hydrophobic Free Energy.** The results reported here reinforce the need to find other approaches to measuring hydrophobic free energy and enthalpy in protein folding. A promising alternative approach has been reported recently: the hydrophobic factor in polymer collapse has been studied with collapsed hydrophobic polymers in aqueous solution by pulling out the polymer using atomic force microscopy and analyzing the force required (31). By studying polymers of different

monomer types, Li and Walker analyzed how hydrophobic free energy contributes to the pulling energetics. The results showed strong correlations with the Kauzmann-Tanford approach to hydrophobic free energy but also showed quantitative differences (31). The pulling free energy values were smaller than expected from solvent transfer studies; the pulling experiments may well contain contributions from pairwise hydrophobic interactions as well as from hydrophobic hydration.

There is a clear analogy between transferring a nonpolar solute out of water into vapor and removing protein nonpolar side chains from water via folding. Lazaridis, et al. (32) examined this factor in the folding energetics by comparing protein unfolding enthalpies converted to vacuum conditions with force-field simulations of protein unfolding in a vacuum. Lazaridis, et al. (32) took calorimetric data for the enthalpy of protein unfolding from Makhatazde and Privalov (33), who converted it to unfolding in vacuum by using empirical calibrations of the heats of desolvating exposed polar and nonpolar groups. This work uncovered a major practical problem in converting experimental unfolding enthalpies to conditions in vacuum: the heats of desolvating the exposed nonpolar and polar groups are large and not known with the accuracy needed. The combined heats of desolvating the exposed polar and nonpolar groups are about 20 times larger than the enthalpy of protein unfolding in water at 25 °C (32).

**Calculating  $\Delta G_{hyd}$  on the Molar (Not Mole Fraction) Concentration Scale.** When calculating  $\Delta G_{LL}$  via two gas-liquid transfers, it is necessary to express the solute concentration on the molar (or number density) scale. Kauzmann (1) recommended calculating  $\Delta G_{LL}$  on the mole fraction scale to avoid including in  $\Delta G_{LL}$  a contribution from the entropy of mixing. There is, however, a basic reason (15–17) for using the molar (or number density) scale when gas-liquid transfer is involved, namely that the large contribution from translational entropy vanishes on this scale. It is intriguing that Ostwald knew this fact (15, 17, 20). Using the mole fraction scale results in including a significant and unwanted contribution from translational entropy (15, 17). Pollack (15) gives the contribution of motional entropy to the chemical potential of a monatomic ideal gas as  $-20.4$  kT, where kT is thermal energy, about  $-0.59$  kcal per mole at 25 °C. The  $\Delta G_{LL}$  values for liquid-liquid transfer of a solute differ significantly between the molar and mole fraction scales (11) and the  $\Delta G_{GL}$  values for gas-liquid transfer differ hugely between the two scales.

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